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## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

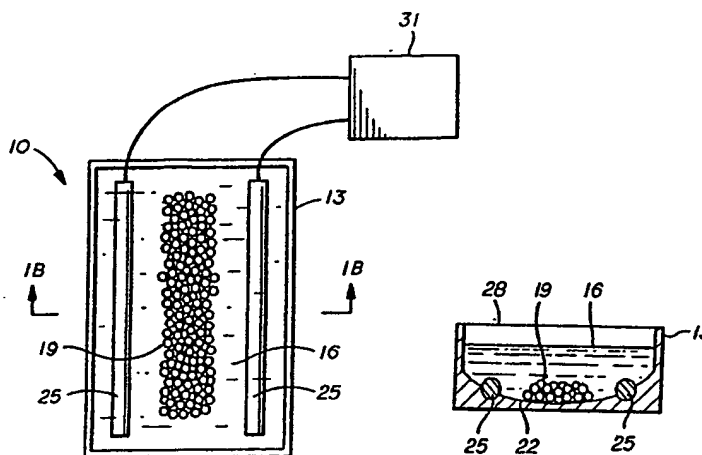
(51) International Patent Classification <sup>4</sup> :  C12N 15/00, 13/00	A2	(11) International Publication Number: WO 89/ 03426  (43) International Publication Date: 20 April 1989 (20.04.89)
<p>(21) International Application Number: PCT/US88/03457</p> <p>(22) International Filing Date: 5 October 1988 (05.10.88)</p> <p>(31) Priority Application Numbers: 106,282 238,607</p> <p>(32) Priority Dates: 9 October 1987 (09.10.87) 30 August 1988 (30.08.88)</p> <p>(33) Priority Country: US</p> <p>(71) Applicant: BAYLOR COLLEGE OF MEDICINE [US/US]; One Baylor Plaza, Houston, TX 77030 (US).</p> <p>(72) Inventor: CHANG, Donald, C. ; 6306 Coachwood, Houston, TX 77035 (US).</p> <p>(74) Agent: PAUL, Thomas, D.; Fulbright &amp; Jaworski, 1301 McKinney St., #5100, Houston, TX 77010 (US).</p>	<p>(81) Designated States: AT (European patent), AU, BE (European patent), CH (European patent), DE (European patent), FR (European patent), GB (European patent), IT (European patent), JP, LU (European patent), NL (European patent), SE (European patent).</p> <p><b>Published</b> <i>Without international search report and to be republished upon receipt of that report.</i></p>	

(54) Title: METHOD OF AND APPARATUS FOR CELL PORATION AND CELL FUSION USING RADIOFREQUENCY ELECTRICAL PULSES

## (57) Abstract

Disclosed are an apparatus and a method for the poration and fusion of cells using high-power radiofrequency electrical pulses. The electrodes of the apparatus can be hand held or part of integrated equipment with special containers for cells. The electrodes, which are positioned equidistant from each other, are attached to a high power function generator. The power function generator can apply a continuous AC electrical field and/or a high-power pulsed radiofrequency electrical field across the electrodes. The alternating electrical field induces cell congregation by the process of dielectrophoresis. The high-power pulsed radiofrequency electrical field porates or fuses the cells.

The method has the ability to porate or fuse biological cells with a very high efficiency. The method can be used to fuse or porate a variety of cells including animal cells, human cells, plant cells, protoplasts, erythrocyte ghosts, liposomes, vesicles, bacteria and yeasts. The method has the unique ability to porate or fuse cells in very small or very large numbers. During the poration or fusions, a variety of chemical agents including DNA, RNA, antibodies, proteins, drugs, molecular probes, hormones, growth factors, enzymes, organic chemicals and inorganic chemicals can be introduced into these cells. The method can also be used to produce new biological species, to make hybridoma cells which produce animal or human monoclonal antibodies and to insert therapeutic genes into human cells which can be transplanted back into the human body to cure genetic diseases.



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METHOD OF AND APPARATUS FOR CELL PORATION AND CELL  
FUSION USING RADIOFREQUENCY ELECTRICAL PULSES

Field of the Invention

This invention relates to the field of poration and fusion of biological cells by application of a high-power pulsed radiofrequency electric field. More particularly, it relates to permeabilizing and fusing cells in a wide variety of fields including gene transfection, micro-injection of cells, production of monoclonal antibodies and making new biological species by hybridization.

Background

Cell poration and cell fusion play a very important role in modern biotechnology. For example, one key procedure in genetic engineering is the introduction of exogeneous genetic material into a host cell. This insertion of genes is accomplished by either permeabilizing the cell membrane to allow entry of genetic material (i.e., gene transfection) or fusing the host cell with a cell containing the desired genetic material. Cell fusion is also important in the production of monoclonal

1 antibodies. The process of producing monoclonal  
antibodies requires the fusion of antibody producing cells  
with continuously dividing cancer cells. (Galfre, G. et  
5 al., Nature 266:550-552 (1977); Lo, M. M. S. et al.,  
Nature 310:794-796 (1984)). Additionally, one highly  
effective method of delivering drugs which normally cannot  
enter a cell is to fuse the cell with liposomes or red  
blood cell ghosts that have been pre-loaded with specific  
10 drugs. (Schlegel & Lieber, Cell Fusion, ed. by A. E.  
Sowers, Plenum Press (1987)).

The conventional techniques of cell fusion rely  
mainly on the actions of viruses (White, J. et al., J.  
Cell Biol. 89:674-679 (1981)); or chemical agents such as  
15 polyethylene glycol (Davidson, R. L. et al., Somatic Cell  
Genetics 2:271-280 (1976)). Virus-induced and  
chemical-induced fusion methods have many shortcomings.  
Not only is the fusion yield often very poor, typically  
less than 0.01%, but the standard fusion techniques may  
20 also cause severe side effects on the fused cells, thus  
greatly limiting their usefulness for many systems.

Alternative methods which induce cell fusion and  
cell poration by electric fields have been developed.  
(Pohl, U. S. Patent No. 4,476,004; Sowers, U. S. Patent  
25 No. 4,622,302; Schoner, U. S. Patent No. 4,578,167;  
Neumann, E. et al. Naturwissenschaften 67:414-415 (1980);  
Zimmerman, U. and Nienken, J., J. Membrane Biol.  
67:165-182 (1982); Bates G. W., et al., Cell Fusion,  
Plenum Press pp. 367-395 (1987)). The basic principle of  
30 these methods of electrofusion is to apply a pulsed high  
strength direct-current (DC) electric field across the  
cell. This DC field is usually generated by briefly  
switching on a DC power source or by discharging a  
capacitor. The applied DC field has a strength of several  
35 kilovolts per centimeter. This external electric field  
induces a large cell membrane potential. When the

1 membrane potential is of sufficient magnitude, a  
reversible breakdown of a small area of the cell membrane  
occurs. The breakdown results in the formation of  
5 physical pores at the surface of the cell. This process  
is called electroporation. Intracellular and  
extracellular material can exchange through the pore while  
it is open. After the DC field is removed, the pore will  
normally reseal quickly. When a pore is created between  
10 two closely adjacent cells a cytoplasmic bridge is formed  
via the pore. When the DC field is turned off the pore  
cannot reseal. Instead, the cytoplasmic bridge usually  
begins to enlarge, eventually causing the two cells to  
fuse.

15       Although the DC electrofusion method has been  
used successfully for a number of biological cells,  
including plant protoplasts (Zimmerman, U. et al.,  
Biochem. Biophys. ACTA 641:160-165.(1981); Bates, G. W. et  
al., Cell Fusion, Plenum Press pp. 479-496 (1987)); blood  
20 erythrocytes (Sowers, A. E., J. Cell. Biol. 102:1358-1362  
(1986); Chang and Hunt, Proceedings of the International  
Symposium on Molecular Mechanisms of Membrane Fusion,  
Buffalo, New York pp. 26 (1987); Stenger, D. A. and Hui,  
S. W., J. Membrane Biol. 93:43-53 (1986)); tumor cells  
25 (Lo, M. M. S. et al., Nature 310:794-796 (1984); Tessie,  
J. et al., Science 216:537-538 (1982)); yeast cells  
(Halfmann, H. J., et al., Archiv. Microbiol. 134:1-4  
(1983)); and blastomeres and eggs (Kubiak, J. Z. and  
Jarkowski, A. K., Exp. Cell Res. 157:561-566 (1985)),  
30 there are still many limitations to the use of this  
method. First, not all cell types can be fused with the  
same ease. In fact many cell types are extremely  
difficult to fuse with DC pulses. Second, there are many  
unknown factors which influence fusion yield. Fusion of  
certain cell types may be successful in one laboratory but  
35 not in others. The DC pulse method is still more of an

1 art than a well understood procedure. Third, it is very  
difficult to use the DC pulse method to fuse cells of  
different sizes. This later problem occurs because the  
5 membrane potential induced by the external DC field is  
proportional to the diameter of the cell. Thus, the  
induced potential is larger for bigger cells. It is  
nearly impossible to chose a proper field strength of  
external field in order to fuse cells of two different  
10 sizes. When the external field is just sufficient to  
cause membrane breakdown in the larger cell, it is  
inadequate to induce a critical membrane potential in the  
smaller cell. On the other hand, if the external field is  
elevated to cause a membrane breakdown in the small cell,  
15 the large potential induced in the larger cell will cause  
an irreversible membrane breakdown and destroy the cell.

The present invention provides an improved method  
of cell poration and cell fusion which overcomes the above  
problems. Unlike the conventional electrofusion method  
20 which employs DC pulses to induce membrane breakdown, the  
present invention uses a pulse or pulses of radiofrequency  
(RF) electric field to reversibly permeabilize cells and  
induce cell fusion. The high-power RF field produces an  
oscillating motion of the cell membrane through a process  
25 of electro-compression. Permeabilization of the cell  
membrane is caused by a combination of electrical  
breakdown and a localized sonication induced from the RF  
field. Thus, this oscillating electric field is more  
effective in breaking down the cell membrane than a DC  
30 field. Since this new method uses only physical means  
(i.e., RF electrical energy) to induce cell poration and  
cell fusion, it is free of biological or chemical  
contamination. The present invention produces results in  
seconds, provides much higher yields than conventional  
35 methods, and has minimal biological side effects. Thus,  
it is a clean, fast, efficient and safe method.

1           The improved efficiency of cell poration and cell  
fusion offered by the method of this invention has a  
particular significance in medical applications. One  
5 example is to produce antibodies for therapeutic uses.  
Since the human body usually rejects animal antibodies,  
such therapeutic antibodies must be produced by hybridomas  
of human cells; however, human hybridomas are extremely  
difficult to form by conventional methods (including  
10 electro-fusion by DC field). The method of the present  
invention will help to improve the efficiency in forming  
human hybridomas. Another example of medical application  
of this method is gene therapy. Many genetic diseases can  
be treated by inserting a therapeutic gene into the  
15 patient's cells in vitro and then transplanting the cells  
back to the patient's body. The conventional methods of  
cell poration (including the DC field method) usually  
require a large number of cells (typically 5-10 million  
cells) to perform a gene transfection and, as a result,  
20 are unsuitable for use in human therapy. In contrast, the  
method of the present invention has been demonstrated to  
be able to transfect cells in small numbers with high  
efficiency, and will be highly useful for gene therapy.

#### Summary of the Invention

25           An object of the present invention is a method  
for the poration of cells.

          An additional object of the present invention is  
a method for the fusion of cells.

30           A further object of the present invention is a  
device for the poration and fusion of cells.

          Another object of the present invention is a  
method for inserting genetic materials into biological  
cells.

35           A further object of the present invention is the  
treatment of genetic disease by inserting therapeutic  
genes into cells that are transplanted into diseased



1 patients.

Another object of the present invention is a method for the formation of hybridoma cells by the fusion  
5 of cells with RF electric field.

An additional object of the present invention is a method which greatly enhances the efficiency of producing monoclonal antibodies.

Another object of the present invention is the  
10 formation of a new species by the fusion of cells from different species using high-power RF pulses.

An additional object of the present invention is the introduction of chemicals and biological molecules into cells by the procedures of poration and/or fusion.

15 Thus, in accomplishing the foregoing objects there is provided in accordance with one aspect of the present invention a method for poration of biological particles comprising the steps of placing a plurality of biological particles in solution between two electrodes  
20 and applying a high-power pulsed RF oscillating field across the electrodes for porating the particles. The biological particles can either be suspended cells in solution or attached cells in cell culture. An additional embodiment of this method includes fusing the biological  
25 particles by placing the suspended biological particles in a container which allows the biological particles to congregate before applying the pulsed RF field.

An alternative method includes fusing the biological particles by applying a low power (e.g., 100 to  
30 400 V/cm) alternating current (AC) electrical field before and/or after the pulsed RF oscillating field. The low-power electric field can cause the particles to move dielectrophoretically to form "pearl chains".

The biological particles can be a variety of  
35 materials including biological cells (human, animal or plant cells), liposomes, vesicles, erythrocyte ghosts,

1 protoplasts, bacteria, and yeasts.

The pulsed RF field applied for the poration and fusion of cells can be an oscillating field of a single frequency or a mixed frequency. The RF oscillating field  
5 may be in the frequency range of 10 KHz to 100 MHz with a pulse width of about 1  $\mu$ sec to 10 msec and a pulse amplitude of up to about 20 KV/cm. In a preferred embodiment the RF oscillating field is about 0.02 to 10  
10 MHz and the pulse width is about 20 to 2000  $\mu$ sec and the pulse amplitude is about 2-10 KV/cm. The wave form of the RF field may be sinusoidal, triangular, sawtooth, or square waves.

Another aspect of the present invention is the fusion of cells for the formation of new species, the  
15 introducing of chemical agents and natural or man-made genetic material into cells, and the formation of hybridoma cells. By the appropriate selection of cell types and materials new species can be formed either by the combining of genetic material from two different  
20 species by the fusion of their cells, or by the isolation or synthesis of the genetic material, and then the introduction of the genetic material into cells by either poration or fusion. Hybridoma cells are made by the fusion of antibody producing cells with continuously  
25 dividing cancer cells. Chemicals, drugs, DNA, RNA and other molecules can be introduced into cells by preloading vesicles, liposomes or erythrocyte ghosts before fusion with target cells.

Another aspect of the present invention is a  
30 device for the poration or fusion of biological particles comprising a container of non-conducting material capable of holding liquid and including an access port for receiving the biological particles. The device also includes electrodes positioned equidistant from each other  
35 and inserted into the container. A high-power function

1 generator is attached to the electrodes and is capable of  
generating a RF electric field and/or an alternating  
electric field. In one embodiment the container is shaped  
5 to allow the biological particles to congregate.

An additional aspect is a device for poration and  
fusion of biological particles comprising a glass chamber  
and used with an optical microscope for observation of the  
poration and fusion of cells.

10 A further aspect is a cell poration and fusion  
device which can be hand-held. This device includes a  
handle and equidistant electrodes. The electrodes can be  
side-attached or bottom-attached and can be designed in a  
variety of shapes including rings, circles, double  
15 helices, squares, ellipses, concentric rings, concentric  
squares, interdigitating arrays, spirals and parallel  
plates.

Other and further objects, features and  
advantages will be apparent from the following description  
20 of the presently preferred embodiment of the invention  
given for the purpose of disclosure when taken in  
conjunction with the accompanying drawings.

#### Brief Description of the Drawings

25 The invention will be more readily understood  
from a reading of the following specification by reference  
to accompanying drawings, forming a part thereof, where  
examples of embodiments of the invention are shown and  
wherein:

30 Figure 1 is a schematic of one form of the  
present invention using a chamber which allows for the  
congregation of cells by gravity. 1A is a top view of the  
device and 1B is a cross-sectional view of the device  
showing the fusion chamber.

35 Figure 2 is a graph of examples of the  
radiofrequency (RF) pulses used in the present invention.

1 2A is a single-frequency symmetrical RF pulse, 2B is an  
asymmetrical RF pulse, 2C is a multiple-frequency RF  
pulse, 2D is consecutive RF pulses of different  
5 frequencies and 2E is a low-power AC field followed by a  
high-power RF pulse followed by a low-power AC field.

Figure 3 is a schematic of one form of the  
present invention showing a large volume chamber for cell  
poration and/or cell fusion. 3A is a top view of the  
10 fusion chamber and 3B is a cross-sectional view showing  
the arrangement of electrodes in the chamber.

Figure 4 is a schematic of a chamber for cell  
poration and/or cell fusion for optical microscopic  
observation. 4A is a elevational view of the chamber and  
15 4B is a cross-sectional view of the chamber.

Figure 5 is a schematic of a hand-held device for  
cell poration and/or cell fusion using a side contact  
configuration. 5A shows an elevational view of the device  
and 5B shows a cross-sectional view of the electrode  
20 inserted inside the cell container.

Figure 6 is a schematic of a double helical  
design for the side-contact electrode assembly. 6A shows  
a elevational view of the helical design for the electrode  
assembly and 6B shows a side view of the same assembly.

25 Figure 7 is a schematic view of a segmented ring  
design for the side-contact electrode assembly. 7A shows  
an elevational view of the electrode assembly, 7B shows  
the connection of the electrode rings in the electrode  
assembly and 7C is a top view of a single electrode ring.

30 Figure 8 is schematic of a rectangular electrode  
assembly for cell poration and cell fusion. 8A shows an  
elevational view of the electrode assembly and 8B shows  
the connection of the electrode squares in the electrode  
assembly.

35 Figure 9 is a schematic of a cell fusion and cell  
poration device with a bottom-contact configuration of

1 electrodes.

Figure 10 is a schematic of the double spiral design for the bottom-contact electrode assembly. 10A shows a elevational view and 10B shows top view of the electrode.

Figure 11 is a schematic view of a concentric ring design for the bottom-contact electrode assembly. 11A shows an elevational view and 11B shows a top view of the electrode.

Figure 12 is a schematic view of different designs for a bottom-contact electrode assembly. 12A is a top view of a square spiral assembly, 12B is a top view of a concentric square assembly, 12C is a top view of an interdigitating array assembly and 12D is a top view of a parallel plate assembly.

Figure 13 is a schematic of a probe for cell poration and cell fusion of a small number of cells using the RF method. The exterior of the metal electrode is designed to fit inside the wells of a 96-well cell culture plate. 13A is a three-dimensional view of the probe, 13B is a cross-sectional view and 13C is a fragmentary elevational view of the electrode.

Figure 14 is a block diagram of the apparatus which provides the source of the AC field for dielectrophoresis and the high-power RF pulses for cell poration and/or cell fusion.

Figure 15 is an electron micrograph showing the surface of a human red blood cell following RF poration treatment. Three RF electric field pulses were applied with a one second interval. The cells were rapidly frozen in liquid freon which was cooled by liquid nitrogen (Temperature 90°K). The frozen sample is examined by freeze-fracture electron microscopy. Magnification 50,000X.

Figure 16 is fluorescent micrographs showing the

1 events of fusion between human red blood cells. Red cells  
were lined up in pearl chains by the process of  
dielectrophoresis. Roughly 10% of the cells were  
5 prelabelled with a fluorescent dye which produced bright  
images under a fluorescence microscope. The unlabelled  
cells could not be seen. 16A shows how the cells looked  
before applying the RF pulses. No transfer of dye between  
labelled and unlabelled cells was seen. 16B shows how the  
10 cells looked 4 minutes after 3 RF pulses (40  $\mu$ sec wide,  
300 KHz, 5 KV/cm) were applied. Some of the labelled  
cells fused with their unlabelled neighboring cells,  
allowing the fluorescent dye to transfer between them.

Figure 17 is a graph showing the measured fusion  
15 yield between human red blood cells using three electrical  
pulses (4 KV/cm, 100  $\mu$ sec). The fusion yield is shown  
to vary with the oscillating frequency.

Figure 18 is a time series of optical micrographs  
showing the fusion of a xanthophore cell with a fish tumor  
20 cell induced by pulsed RF fields. 15A is before fusion  
but after the xanthophore (marked by the arrow) was  
brought into close contact with two tumor cells by  
dielectrophoresis. 15B is two minutes after application  
of the RF pulses showing that the xanthophore has already  
25 begun fusing with one of the tumor cells. 15C is 4  
minutes after application of the RF pulses showing that  
the xanthophore and tumor cell have completely merged into  
a single round cell.

#### DETAILED DESCRIPTION

30 In the description which follows, like parts are  
marked throughout the specifications and drawings with the  
same referenced numerals. The drawings are not  
necessarily to scale and certain features of the invention  
may be exaggerated in scale or shown in schematic form in  
the interest of clarity and conciseness. It will be  
35 readily apparent to one skilled in the art that various

1 substitutions and modifications may be made to the invention disclosed herein without departing from the scope and spirit of the invention.

5 One embodiment comprises a method for poration of biological particles comprising the steps of placing the biological particles in solution between two electrodes and applying a pulsed radiofrequency (RF) oscillating electric field across the electrodes, Fig. 1. A variety  
10 of biological particles can be used including biological cells, erythrocyte ghosts, liposomes, protoplasts, bacteria and yeasts. The biological particles can be suspended cells in solution or can be attached cells in cell culture.

15 When a cell is placed in an electric field, an electrical potential is induced across the cell membrane. For a spherical cell, the membrane potential induced by an external electric field is

$$V_m = 1.5 r E \cos \theta \quad (1)$$

20 where  $r$  is the radius of the cell,  $E$  is the strength of the external field and  $\theta$  is the angle between the direction of the external field and the normal vector of the membrane at the specific site.

The induced electric field within the membrane is

25  $E_m = V_m/d = 1.5 (r/d) E \cos \theta \quad (2)$

where  $d$  is the thickness of the membrane. Since  $d$  is much smaller than  $r$  ( $d$  is about  $6 \times 10^{-7}$  cm while  $r$  is in the order of several microns),  $E_m$  is about 1000 fold larger than the applied field,  $E$ . The large electric  
30 field within the membrane produces two effects. First, it exerts a strong force on the phosphate head group of the lipid molecules in the membrane and tends to move them in the direction of the field. Secondly, it compresses the membrane. When the external electric field oscillates,  
35 the lipid molecules within the membrane also undergo an oscillating motion.

1 In this arrangement, the cell itself functions as  
an antenna and the membrane is a transducer which converts  
the electrical oscillation into a mechanical oscillation.  
5 Thus, it is possible to generate an ultrasonic motion in  
the cell membrane by applying an external RF field.  
Because the induced potential at a given site of the  
membrane is a function of the angle between the  
orientation of the membrane and the electric field vector,  
10 the induced potential is not uniform over the entire cell  
surface. The applied energy is focused at the poles of  
the cell, that is, at  $\theta = 0^\circ$  or  $180^\circ$ . Because the  
amplitude of the external field can be adjusted such that  
there is sufficient sonication power to break down the  
15 cell membrane at the poles but not at other parts of the  
membrane, the sonication can be localized. Experiments  
indicated that this localized membrane breakdown induced  
by the externally applied pulsed RF field is reversible.  
That is, the pore(s) induced by the RF field reseal  
20 quickly (within minutes) after the field is turned off.  
Furthermore, most of the cells apparently stay viable.

Such temporary permeabilization of the cell  
membrane is called cell poration. During this time period  
when pores are formed, a brief exchange of intracellular  
25 and extracellular materials occur. Many molecules,  
including drugs, antibodies, and gene segments, which  
normally cannot penetrate the cell membrane, can enter the  
cell through the temporarily opened pores that were  
induced by the pulsed RF field.

30 Another embodiment of this invention comprises a  
method for fusing cells. In order for biological  
particles to be fused, they must be in close proximity.  
When cells are in close proximity they are said to  
congregate. Two alternative procedures may be used to  
congregate the cells before fusion. In one, a container  
35 with a shape that allows the biological particles to



1  
congregate by gravity is used. For example, the bottom of  
the container can be made in a concave shape (see Figs. 1  
and 3). This allows the cells to congregate. When the  
5 cell membranes are permeabilized by the applied RF field,  
the closely adjacent cells can form cytoplasmic bridges.  
This process results in the fusion of cells.

Alternatively, a low amplitude continuous  
alternating current (AC) electrical field can be applied  
10 across the two electrodes. The frequency ranges from  
about 60 Hz to about 10 mega Hz. Typically a 100-400 V/cm  
field strength is used. Under the low amplitude AC field  
the cells act as dipoles and line up parallel to the  
field, eventually forming a long chain of cells which  
15 appear like "pearl chains". This process is called  
"dielectrophoresis" (Schwan, H. P. and Sher, L. D., J.  
Electrochem. Soc. 116:22C-26C (1969); Pohl, H. A. et al.,  
J. Biol. Phys. 9:67-86 (1981)). Formation of this pearl  
chain normally takes about a few seconds to one minute.

20 The present invention uses a pulsed RF field to  
porate and/or fuse cells and has a clear advantage over  
the conventional electro-fusion method that uses a pulsed  
DC field. First, the RF field is a much more efficient  
means of transmitting energy to the cell membrane than the  
25 direct current field. The present invention utilizes a  
localized sonication to break down the cell membrane.  
This method is much more effective than the DC pulse  
method which relies solely on the electrical breakdown.  
The cell membrane is composed of macromolecules which have  
30 characteristic frequencies of thermal motion. When the  
frequency of the applied oscillating field matches one of  
these natural frequencies, a condition of resonance is  
reached, and the efficiency of energy transfer is greatly  
enhanced. In real biological cells the resonance peak can  
35 be very broad. The pulsed radiofrequency field can be  
carefully varied to achieve the proper resonant frequency

1 for the cells of interest. Consequently, the ability to  
induce membrane breakdown will require less power than  
using a direct current field and results in less risk of  
5 irreversibly damaging the cell.

Second, this invention overcomes the difficulties  
encountered when the conventional methods are used to fuse  
cells of different size. In order to produce an  
electrical breakdown of the cell membrane, the  
10 field-induced membrane potential must exceed a certain  
critical value,  $V_c$  (typically 1 volt). Such breakdown  
can be reversible, and the membrane will reseal after the  
external field is turned off if the induced membrane  
potential is not too much larger than  $V_c$ . The cell  
15 normally remains viable after such reversible breakdown.  
On the other hand, if the induced potential is much higher  
than  $V_c$ , the membrane breakdown is irreversible, the  
cell is permanently damaged, and will not remain viable.

From Eq. 1 it can be seen that when cells of  
20 different sizes are placed inside an electric field, the  
induced membrane potential is higher for the larger cell  
than for the smaller cell. This size-dependence of  
membrane potential causes a problem when attempting to  
fuse cells of different sizes using a DC field. Assume  
25 that two cells, A and B, are to be fused and that the  
radius of cell A,  $r_a$ , is about twice as big as the  
radius of cell B,  $r_b$ . In order to cause a reversible  
membrane breakdown in cell B, the applied external field  
must be sufficient so that  $1.5 E r_b$  is greater than  
30  $V_c$ . However, the same applied electric field will  
induce a much larger  $V_m$  in Cell A, and will cause an  
irreversible breakdown of the membrane leading to damage  
to this cell. Thus it is very difficult to use direct  
current pulses to fuse cells of significantly different  
35 sizes.

This problem can be solved by applying a pulsed

1 radiofrequency field. When the applied field is a  
radiofrequency oscillating field instead of a DC field,  
the amplitude of the induced membrane potential is a  
5 function of the frequency. The membrane potential  
predicted in Eq. (1) is derived under the steady state  
condition. The induced potential does not arise  
instantaneously upon the application of the external  
field. If the external field is stationary, the membrane  
10 potential will reach  $V_m$  given a sufficient time. The  
time required to establish this steady state membrane  
potential is called "relaxation time", or  $\tau$ , which is  
given by

$$1/\tau = 1/R_m C_m + 1/r C_m (R_i + 0.5R_e) \quad (3)$$

15 where  $R_m$  and  $C_m$  are specific resistance and specific  
capacitance of the membrane, and  $R_i$  and  $R_e$  are the  
specific resistances of the intracellular medium and the  
extracellular medium, respectively. (C. Holzapfel et al.,  
J. Membrane Biol., 67:13-26 (1982)). For a cell of  
20 several microns in diameter,  $\tau$  is typically in the order  
of 1  $\mu$ sec.

Since  $R_m$  in most cells is very large, for  
practical purposes, eq. (3) can be simplified to

$$\tau = r C_m (R_i + 0.5R_e) \quad (4)$$

25 Thus the relaxation time is approximately proportional to  
the radius of the cell.

Because the build-up of the membrane potential  
requires a time period characterized by the relaxation  
time  $\tau$ , the membrane potential induced by a RF field is  
30 frequency dependent. If a radiofrequency field is applied  
at a frequency smaller than  $1/\tau$ , the membrane potential  
has no problem in following the external field. The  
applied field will produce a 100% cellular response in  
 $V_m$ . On the other hand, if the frequency of the applied  
35 radiofrequency field is greater than  $1/\tau$ , the membrane  
potential cannot catch up with the changes in the applied

1 field, and the response of the membrane potential will be  
less than 100%. In general, the maximum membrane  
potential induced by a RF field is

$$V(\omega) = 1.5 rE \cos \theta X(\omega) \quad (5)$$

5 where  $r$ ,  $E$  and  $\theta$  have the same meaning as in Eq. (1),  
 $\omega$  is the angular frequency, and  $X(\omega)$  is a function of  
the frequency such that

$$X(\omega) = [1 + (\omega\tau)^2]^{-1/2} \quad (6)$$

10 when  $\omega < 1/\tau$ ,  $X(\omega)$  is near unity.  
When  $\omega > 1/\tau$ ,  $X(\omega)$  decreases very rapidly with  
increasing frequency.

This frequency dependent effect can be used to  
15 fuse cells of different sizes. From Eq. (4),  $\tau$  of the  
cell is roughly proportional to  $r$ . Thus the larger cell  
will have a longer  $\tau$ . To fuse the A and B cells, a  
pulsed RF electric field that has a frequency  $\omega$  is  
applied such that

$$1/\tau_a < \omega < 1/\tau_b \quad (7)$$

20 Since the frequency is less than  $1/\tau_b$ ,  $X(\omega)$   
approaches unity for cell B and thus the field will  
produce a full effect on the small cell. On the other  
hand, since the frequency is greater than  $1/\tau_a$ , the  
25 induced membrane potential in cell A cannot fully follow  
the variation of the applied field, that is,  $X(\omega)$  in  
cell A is less than unity. Thus, in a pulsed  
radiofrequency field, the effect of the stimulating field  
sensed by the small cell is greater than the effect on the  
30 large cell. Consequently, a pulsed radiofrequency field  
can be applied which induces a reversible breakdown of the  
membrane of the small cell without irreversibly damaging  
the larger cell.

One embodiment of a device 10 for poration and/or  
35 fusion of biological particles is shown in Figure 1. It  
is a fusion chamber which includes a non-conducting

1 container 13 for holding the solution 16 of biological  
particles 19. The container has a slightly concave bottom  
22 so that biological particles 19 will congregate, under  
5 gravity, between the electrodes 25. The electrodes 25 are  
a pair of equidistant metal wires or metal bands made of  
nontoxic material, such as platinum or surgical stainless  
steel. The electrodes can be parallel wires or can be in  
almost any shape or design. The container 13 has an  
10 access port 28 wherein biological particles 19 can be  
added or removed.

To induce cell-poration or cell-fusion, a high  
power function generator 31 generates one or many high  
power RF pulses which are applied through the pair of  
15 electrodes 25. The pulse shape can include one of those  
shown in Fig. 2. In Fig. 2A, the pulse is a symmetrical  
RF oscillation with a single frequency. In Fig. 2B, the  
RF pulse consists of a single frequency asymmetrical  
sinusoidal wave. In Fig. 2C, the RF pulse contains a  
20 mixture of sinusoidal waves of multiple frequencies (in  
this example, two frequencies). In Fig. 2D, alternating  
sinusoidal pulses of different frequency are used. In the  
preferred embodiment, the pulse shown in Fig. 2B is used,  
because it allows the applied energy of the field to be  
25 used more efficiently in inducing cell poration or  
fusion. Although the preferred RF electric field wave  
form is sinusoidal, other wave forms with repetitive  
shapes can be used. For example, triangular waves, square  
waves and sawtooth waves can be used to fuse or porate  
30 cells of different types.

One skilled in the art will readily recognize  
that the parameters of the pulsed field are changed to  
accommodate the characteristics of the different  
biological samples. The radiofrequency within the pulse  
35 may vary over the complete radiofrequency range of 10 KHz  
to 100 MHz. Typically a value in the order of 0.02 to 10

1 MHz is used for the poration and/or fusion of biological cells.

The width of the pulse may vary from about 1  $\mu$ sec to 10 msec. In the preferred embodiment approximately 20 $\mu$ sec to 2 msec is used.

The field strength is controlled by varying the pulse amplitude. For fusion and poration of cells the range of 1 to 20 kV/cm is employed. In the preferred embodiment pulses of field strength up to about 10 KV/cm are used.

The pulse can be a single pulse, a train of pulses or multiple trains of pulses. A train of pulses are multiple pulses with an interval in between; for example, a series of ten pulses 0.5 milliseconds in width each pulse separated by 0.5 seconds. In some instances such as the fusion of HL-60 cells, the maximum fusion yield is enhanced by applying multiple pulses.

The RF pulses used for cell-poration and cell-fusion are similar. The main difference is that in cell fusion, the cells need to congregate (be brought into close proximity) before the high power RF pulse is applied. Furthermore, the cells must be maintained in close proximity after application of the RF pulse. The above described device brought the cells together by gravitational congregation. An alternative, and more efficient method of cell aggregation is dielectrophoresis, where a continuous alternating current (AC) electric field is applied across the electrodes before and/or after the application of the high-power RF pulse. The amplitude of this continuous AC field is typically in the range of 100 to 400 V/cm. Its frequency may vary from about 60 Hz to about 10 MHz. During cell fusion in the preferred embodiment the actual electric field applied across the electrodes may look like that shown in Fig. 2E.

Another device for poration and/or fusion of

1 larger volumes of cells is shown in Fig. 3. An array of  
equidistant electrodes 25 instead of a single pair of  
electrodes is used to apply the AC field and the pulsed RF  
5 field. The bottom of this fusion chamber can be either  
flat or slightly concave. It is made of transparent  
material such as glass or clear plastic. This chamber can  
be placed on top of an inverted optical microscope so that  
the events of cell fusion and/or cell poration can be  
10 directly monitored. Since the effects of different  
experimental conditions can be assayed in a timely manner  
with the design, it will be useful for establishing the  
optimal condition for cell fusion and/or cell poration.

The electrodes can be arranged in any pattern, as  
15 long as they are maintained equidistant from each other.  
In the preferred embodiments the patterns have included  
interdigitating array, concentric circles and double  
spirals.

Another preferred device 10 for cell poration and  
20 cell fusion is shown in Fig. 4. This device 10 is  
designed to allow observation of cell fusion under an  
optical microscope using a small volume of cell  
suspension. This device is formed by two glass plates 34  
separated by spacers 37 of approximately 0.3 mm thickness,  
25 with the cell suspension 19 sandwiched between the glass  
plates 34. In one embodiment thin glass plates such as  
cover slips are used. Electrodes 25 are two parallel  
platinum wires which are about 0.5 mm apart. The platinum  
wire electrodes 25 are connected to a high-power function  
30 generator 31. The high-power function generator can  
generate both alternating current electric fields and  
pulsed radiofrequency fields. An inlet tubing 41 and an  
outlet tubing 44 are used to insert and remove cells from  
the space between the electrodes.

35 Another embodiment of the present invention for  
cell poration and cell fusion is shown in Fig. 5. The

1 purpose of this device is to porate or fuse a very large  
volume of suspended biological particles; including  
biological cells, protoplasts, bacteria and yeasts. This  
5 device 20 is designed for ease in application,  
maintenance, and cleaning. The cell suspension is  
contained in a non-conducting cylindrical container 13.  
The electrode assembly 50 is attached to an insulating  
handle 47. To porate or fuse the suspended cells, the  
10 electrode assembly is lowered into the cell container 13  
by manipulating the handle 47. The electrodes 25 are  
connected to the high-power function generator 31 by a  
connection means 49. The AC field for cell fusion and the  
high power RF pulses for cell poration and/or cell fusion  
15 are then applied through the electrodes 25 in the  
electrode assembly 50.

In this device 20 the electrode assembly 50 is a  
vertical cylinder 53 and metal electrodes 25 are exposed  
at the side (i.e., the cylindrical surface). The cylinder  
20 can be any non-conducting material, for example, glass,  
plastic, or teflon. When the electrode assembly 50 is  
lowered into the cell container 13, the suspended cells 19  
are displaced and form a thin layer of cell suspension 19  
surrounding the electrode assembly 50. Thus, all cells  
25 are in close proximity of the electrodes. When an  
electrical potential is applied across the electrodes, the  
cells are exposed to the electric field.

One design of the electrode assembly 50 is shown  
in Fig. 6. Two metal wires or bands are coiled to form a  
30 double helix electrode 25. The helices are identical in  
shape except one is positioned between the other. These  
two helices are attached to a cylindrical support 53. The  
spacing between these two helices 25 is kept constant.  
Thus, when an electrical potential is applied across the  
two metal wires, the amplitude of the electric field  
35 generated between the two helices is uniform along their



1 entire length.

Another embodiment of the electrode assembly 50 for cell poration and cell fusion is shown in Fig. 7.  
5 Here the electrode 50 assembly is comprised of a stack of metal ring electrodes 25 separated by non-conducting insulating spacers 53 of fixed thickness. These ring electrodes 25 are connected together in an alternating fashion to form two sets of electrodes 25, each of which  
10 is then connected to the output terminals of the high-power function generator. The rings have an attachment means 56 and a hollow area 59 for the passage of the wire to the alternate electrode 25.

The electrodes 25 do not have to be circular, but  
15 can be any shape. Shapes which can be used include circular, rectangular as in Fig. 8 or elliptical.

Another embodiment for cell poration and cell fusion is shown in Fig. 9. The cell suspension 19 is contained in a non-conductive container 13. An electrode  
20 assembly 50 is attached to a handle 47 which can be used to manipulate the position of the electrodes. Unlike the previous devices, the electrodes of this embodiment are exposed at the bottom of the electrode assembly 50. This device is thus particularly useful in porating and/or  
25 fusing cultured cells that attach to the bottom of culture dishes.

One design of the bottom-contact electrode assembly 50 is shown in Fig. 10. The electrode assembly 50 consists of two spirals of metal bands, which serve as  
30 the "ground" (-) and "high voltage" (+) electrodes 25. The two spirals are positioned in such a way that the spacing between each spiral is maintained constant. The equal spacing arrangement ensures that an applied electric field across the two electrodes 25 is uniform in strength  
35 throughout the entire area covered by the electrode assembly.

1 In addition to the spiral design, other configurations including, multiple concentric rings, rectangular shapes, interdigitating arrays, parallel  
5 plates or elliptical shapes can be used (see Figs. 11 and 12). The rings or shapes connected in alternating fashion into two groups. One group of these rings or shapes is connected to the "ground" (-) terminal, while the other group of rings or shapes are connected to the "high  
10 voltage" (+) terminal of the high-power function generator. The spacing between the rings or shapes is constant so that the strength of the electric field generated between the adjacent rings or shapes is uniform throughout the entire assembly. In the bottom-contact  
15 electrode assemblies, the electrodes can be wires, plates or bands. In the preferred embodiment, the width of the electrodes is greater than the depth of the cell suspension.

Another embodiment of the present invention for  
20 cell poration and cell fusion is shown in Figs. 13A-C. The probe 20 allows cell fusion or gene transfection for a small volume of cell suspension. The probe 20 will fit into a flat-bottomed 96-well cell culture plate, for example Corning model 25860. The probe 20 includes two  
25 coaxial electrodes 25. The inner electrode 25a is a solid cylinder and the outer electrode 25b is a hollow tube. The coaxial electrodes 25 can be made of a variety of conductive materials. In the preferred embodiment, the coaxial electrodes 25 are made of stainless steel. The  
30 coaxial electrodes 25 are attached to a nonconductive insulating holder 54 preferably made of teflon or plastic.

The gap between the inner 25a and outer 25b coaxial electrodes may vary from about 0.5 to 2.0 mm. In  
a preferred embodiment, the electrode 25 has a 0.7 mm  
35 gap. With this probe 20 the total volume of suspended cells to be fused or porated is about 80  $\mu$ l and it is

1 possible to do cell fusion or cell poration with as little  
as 20  $\mu$ l of cell suspension.

The probe 20 has a handle 47 made of  
5 non-conductive material, preferably teflon. Holding means  
55, hold the outer electrode 25a in place.

This design has several advantages. Besides  
allowing the use of small volumes of cell suspension for  
cell fusion or cell poration, it is also simple to use and  
10 highly cost-effective. Unlike most commercial machines  
which require one cuvette to transfect one cell sample,  
this probe can serially transfect many cell samples using  
plates with multiple wells.

Another embodiment of the present invention for  
15 cell poration and cell fusion is shown in Figure 14. This  
figure shows the block diagram of the high power function  
generator which generates both the AC field for  
dielectrophoresis and the high power RF pulses for cell  
fusion and/or cell poration. The switching between the AC  
20 field and the RF field is controlled by a mercury wetted  
relay. The RF pulses are generated by gating the output  
of a radiofrequency oscillator and then passing through a  
MOSFET power amplifier, the power output of which may be  
as high as twenty kilowatts.

25 Alternatively the AC field and the pulsed RF  
field can be generated by synthesizing the required  
electrical wave with a digital computer and amplifying  
these wave forms using a power amplifier. In this  
embodiment the protocol can be controlled entirely by the  
30 computer and thus no switching relay is needed. This  
computer-synthesized high power function generator has  
several advantages. First, very complicated wave forms  
can be generated to optimize the fusion and/or poration of  
different types of cells. Second, when the high power  
function generator is used in more than one protocol or by  
35 more than one user, each protocol can be stored separately

1 in a data storage device for example, a magnetic  
diskette. Since the protocols can be recalled quickly,  
the high power function generator can be reprogrammed to  
5 generate the desired wave forms without manually  
readjusting all the parameters. Third, the same computer  
can be used as a digital oscilloscope to record the actual  
electrical field applied to the cells. This record can be  
saved in a data storage device as the permanent record of  
10 any particular cell fusion or cell poration experiment.

Excessive current is harmful to the cell because  
of the resulting thermal effects and pH changes. To avoid  
generating excessive current and the resulting effects  
during the application of the electric field, the  
15 suspension medium of the cells is usually a low ionic  
strength solution. Preferably it contains very low  
concentration of salts. A typical suspension medium may  
contain 1 mM of electrolyte including 0.4 mM Mg-acetate  
and 0.1 mM Ca-acetate. The medium is buffered and the pH  
20 maintained in the physiological range, for example, pH  
7.5. Any buffer commonly used for biological purposes,  
for example, 1 mM HEPES

(N-2-hydroxyethyl-piperazine-N'-2-ethane sulfonic acid) is  
adequate for cell poration and/or cell fusion.  
25 Non-electrolytes are added to maintain the osmolarity of  
the medium at about the osmolarity of extracellular  
fluid. In the preferred embodiment, relatively high  
molecular weight, cell impermeable carbohydrates, such as  
sucrose and mannitol, are used to maintain the osmolarity.

30 For some cells, a slightly higher ionic strength  
in the medium seems to improve the fusion yield. For  
example, human erythrocytes fuse easily in 30mM  
Na-phosphate. Thus, the present method of fusion can use  
suspension medium with an ionic strength ranging from 0.1  
mM to 100mM depending on the cell type.

35 The present invention for cell poration and cell

1 fusion has a variety of uses. Many biological active  
substances, including DNA, RNA, organic chemicals,  
inorganic chemicals, drugs, antibodies, proteins,  
5 hormones, growth factors, enzymes and radio- or  
fluorescent-labelled molecular probes normally cannot be  
readily taken up by cells. The present invention provides  
an effective method to transport these biological active  
substances into the cells. In one embodiment of the  
10 present invention, cells can be temporarily permeabilized,  
that is porated, by applying high-power RF pulses and the  
biological active substances can then enter the cells  
during this poration period. The porated cells can be  
biological cells (including, animal, human or plant  
15 cells), protoplasts, bacteria or yeasts. In another  
embodiment of the present invention, the biological active  
substances can be inserted into the cells by fusing the  
target cells with other biological particles which have  
been pre-loaded with the active substances. Such  
20 biological particles include liposomes and erythrocyte  
ghosts, which can be easily preloaded with desired  
substances using a standard osmotic shock and dialysis  
method. (Schlegel & Lieber Cell Fusion ed by A.E. Sowers  
Plenum Press (1987)). The target cells may be any cells  
25 which will receive the biological active substances and  
include isolated cells, egg cells, embryonic cells, any  
primary or transformed cultured cells, or other cells in  
vitro.

In like manner, biological substances could be  
30 extracted from biological cells. For example, many  
molecules such as hormones, growth factors, enzymes,  
proteins and nucleic acids may not be able to cross the  
membrane barrier. Using the poration method of the  
present invention, temporary pores can be induced in the  
cell membrane. The non-permeable molecules can then exit  
35 the cell. This procedure could be useful in a variety of

1 industries which use growing cells to produce biological  
molecules. This procedure allows the extraction of these  
molecules without having to kill the cells.

5 Example I

The ability and efficiency of the RF  
electroporation method to insert foreign genes into the  
target cell is examined using the cultured eukaryotic  
fibroblast cell line COS-M6 (M6). Chloramphenicol  
10 AcetylTransferase (CAT) DNA was used as a gene marker.  
Bacterial CAT DNA was inserted into a plasmid vector  
(pSV<sub>2</sub>-CAT). The CAT enzyme is not endogenously produced  
in mammalian cells, such as M6. Thus, the amount of CAT  
gene incorporated into the target cells can be assayed by  
15 monitoring the amount of CAT enzyme produced after the  
transfection.

The protocol was to apply 3 trains of high-power  
RF pulses at 10 sec intervals. Each train consists of 5  
pulses (frequency 100 KHz, field strength 2.5 KV/cm, pulse  
20 width 0.5 msec).

The RF poration protocol of the present invention  
is a highly effective method of gene transfection. In the  
conventional methods of gene transfection, for example,  
the calcium phosphate method or the DEAE-dextran method,  
25 usually requires at least 5-10 µg of plasmid DNA for  
each transfection. In previous electroporation methods  
that used DC pulses, even larger amounts of DNA (typically  
10-40 µg) were required. (Ansubel et al., Current  
Protocols in Molecular Biology, John Wiley & Sons, 1988).  
30 Using the RF poration method of this invention, we  
obtained a high level of CAT activity (76% acetylation per  
25 µg of protein) when M6 cells were transfected using  
only 0.1 µg of CAT DNA. Furthermore, up to 10.6%  
acetylation per 25 µg of protein was observed when M6  
35 cells were transfected with as little as 0.01µg  
of CAT DNA. Thus, it is evident that the RF poration

1 method has a much higher efficiency of gene transfection.  
The improved efficiency not only results in great savings  
in labor and material that is required to produce DNA, but  
5 also will allow the transfection of cells which were  
previously difficult to transfect.

Another advantage of the RF poration method is  
that it requires far less cells for gene transfection.  
The conventional chemical methods and the DC  
10 electroporation method typically require 2 to 10 million  
cells to do one transfection. With the RF method, M6  
cells have been transfected with the CAT gene in high  
efficiency using as few as 0.1 million cells. Further  
experiments indicated that even lower numbers of cells ( $1$   
15  $\times 10^4$ ) can be used. Currently, the minimum cell number  
is limited by the amount of total cell protein required to  
perform the CAT assay and not the ability to transfect  
cells. (Typically 25 micrograms of total cellular protein  
are needed for the CAT reaction.)

#### 20 Example II

Because of the unique abilities of the RF  
poration method to transfect cells in small quantity and  
with high efficiency, the method will be particularly  
useful in the development of gene therapy. Many diseases  
25 are known to be caused by genetic defects. Such diseases  
could be treated by inserting a therapeutic gene into  
human cells such as bone marrow stem cells and then  
transplanting these cells into the human body.

For example, patients with sickle cell anemia  
30 have a defective gene which produces abnormal hemoglobin.  
To treat such a genetic disease, bone marrow stem cells  
are extracted from the patient and transfected with the  
normal hemoglobin gene. The transfected stem cells are  
transplanted back into the patient. With the appropriate  
35 vector the normal gene will be stably integrated into the  
genome and the patient will be able to produce normal

1 hemoglobin.

The key step in this treatment is the transfection of the bone marrow stem cells with the normal gene. Because the number of stem cells which are extracted is relatively small, a gene transfection method of high efficiency that is suitable for extremely low cell numbers is required. The method of the present invention of poration using RF pulses uniquely has this ability. Thus this method will be highly useful for gene therapy.

The usefulness of this method for gene therapy is not limited to sickle cell anemia. This method can be applied to insert normal genes into human cells to cure many genetic diseases. Other examples include: introducing the gene for clotting factor VIII into bone marrow stem cells to cure hemophiliacs; inserting the gene for insulin into pancreatic islet cells or other human cells to treat diabetes; introducing the gene for the human LDL (low density lipoprotein) receptor into liver cells or other human cells to lower the cholesterol level in the bloodstream of hypercholesterolemia patients; and introducing the gene for human growth hormone into human cells to correct growth defects. Thus, the possibilities of using this method to insert genes into human cells to treat genetic diseases is unlimited.

#### Example III

#### Morphological Changes of the Cell Membrane during the Process of RF Field Electroporation

A fraction of a second after human red blood cells were exposed to RF pulses, they were rapidly frozen in liquid freon cooled by liquid nitrogen. The structure of the cell membranes were examined using the technique of freeze-fracture electron microscopy. In Fig. 14 the electron micrograph shows the surface structure of the red blood cell after 3 RF pulses (400 kHz, 40  $\mu$ sec wide, 5



1 kV/cm field strength) were applied. Membrane pores with  
diameters of 0.1 to 0.3 micrometers were clearly seen.  
These pores are sufficiently large to allow a large piece  
5 of DNA to easily diffuse from the extracellular medium  
into the cell. Thus, there is direct evidence that the  
applied RF fields can induce large pores at the cell  
surface. The morphological evidence clearly shows that  
the method of the present invention is effective in  
10 inducing membrane poration to allow transfection of cells  
with exogeneous genes.

#### Example IV

An example of the advantage that the present  
invention has over the conventional DC (direct current)  
15 electrofusion method was seen in the fusion of human  
erythrocytes. The fusion events were assayed by labelling  
the membranes of a small number of the suspended cells  
with a lipophyllic fluorescent dye, for example,  
1,1',-dihexadecyl-3,3,3",3'-tetramethylendocarbocyanine  
20 perchlorate. The cells were observed with a fluorescence  
microscope. Before applying the RF pulses, only the  
prelabelled cells give a fluorescent image and they  
appeared as isolated cells (see Figure 16A). After the  
cells were exposed to pulsed RF fields, unlabelled cells  
25 started to fuse with labelled cells and the dye was  
gradually transferred from the labelled cell to the  
unlabelled cell. Eventually both cells became labelled  
(see Figure 16B). This fusion process took only a few  
minutes following the application of the RF pulses.

30 Two types of cell fusion were observed in this  
experiment: (1) Membrane fusion, in which the fluorescent  
dye was transferred from the labelled cell to the  
unlabelled cell but the two cells did not merge their  
cytoplasm; and (2) cytoplasmic fusion, in which the fusing  
35 cells merged together to form one single large cell. The  
percentage of cells undergoing cytoplasmic fusion depends

1 strongly on the oscillating frequency of the applied RF  
field. The fusion yield for erythrocytes after RF pulses  
of different frequency are applied is shown in Figure 17.  
5 The highest yield of fused cells occurred when the applied  
RF field was oscillating at 100KHz. The fusion yield  
decreased to a very low level as the frequency became too  
high or too low. No cytoplasmic fusion was detected when  
the applied field was in the form of DC pulses with the  
10 same pulse amplitude and pulse width as the RF pulses.  
These results clearly indicate that the RF pulse method of  
this invention is much more effective in inducing cell  
fusion than the DC pulse method.

Another example of the advantage of the present  
15 invention over the DC electrofusion method is in the  
fusion of human erythrocytes with a human leukemia  
cultured cell line, HL-60. Fusion of these two cells  
types was not obtainable using the DC pulse method. The  
failure is probably due to the differences in cell size;  
erythrocytes are significantly smaller than HL-60 cells.  
20 However, using the fluorescent dye assay and the pulsed RF  
field of the present invention, we were able to obtain the  
fusion of erythrocytes with HL-60 cells.

#### Example V

25 The RF pulse method can be used to fuse cells to  
make hybridomas. Pigment cells from goldfish  
(xanthophores) were fused with a tumor cell line derived  
from fish skin cells. Because xanthophore cells have a  
built-in histochemical marker (the carotenoid droplets),  
30 it is comparatively easy to assay their fusion with  
non-pigmented tumor cells. Fig. 18 shows the sequential  
steps in the fusion of a xanthophore and a skin tumor  
cell. In Fig. 18A the cells were brought into close  
contact by dielectrophoresis. Three pulses of RF field  
35 (40  $\mu$ sec wide, frequency 400 kHz, field strength 3.3  
kV/cm) were then applied. Within two minutes the

1 cytoplasms of the two cells begun to merge (see Figure  
18B). After 4 minutes, the cells completely coalesced  
into a single giant cell (see Figure 18C).

5 An important application of forming hybridomas  
using the RF pulse method is to make antibodies,  
especially human monoclonal antibodies. In this instance  
the biological particles to be fused can include antibody  
producing cells (for example, lymphocyte B cells) and  
10 continuously dividing cells (for example, cancer cells).  
Using a selection process, the resultant hybridoma cells  
can be cultured to produce specific monoclonal  
antibodies.

One skilled in the art will readily appreciate  
15 the present invention is well adapted to carry out the  
objects and obtain the ends and advantages mentioned, as  
well as those inherent therein. The devices, methods,  
procedures and techniques described herein are presently  
representative of the preferred embodiments, are intended  
20 to be exemplary, and are not intended as limitations of  
the scope. Changes therein and other uses will occur to  
those skilled in the art which are encompassed within the  
spirit of the invention or defined by the scope of the  
appended claims.

25 What is claimed is:

30

35

1

CLAIMS

1. A method for poration or permeabilization of  
5 biological particles, comprising the steps of:

placing a plurality of biological particles  
between electrodes; and

applying a pulsed radiofrequency oscillating  
electrical field across said electrodes.

10

2. The method of Claim 1, wherein said  
biological particles are selected from the group  
consisting of animal cells, human cells, plant cells,  
protoplasts, bacteria and yeasts.

15

3. The method of Claim 1, for injecting  
chemical agents or biologically active molecules into  
biological cells, wherein said chemical agents or  
biologically active molecules are selected from the group  
20 consisting of DNA, RNA, antibodies, proteins, drugs,  
hormones, growth factors, enzymes, organic chemicals, and  
inorganic chemicals.

4. The method of Claim 1, for extracting  
25 molecules from biological cells, wherein said molecules  
are selected from the group consisting of proteins,  
nucleic acids, hormones, growth factors, enzymes, and  
other biologically active molecules.

5. The method of Claim 1, wherein said pulsed  
30 radiofrequency field includes a frequency range of about  
10 KHz to 100 MHz, a pulse width range of about 1  $\mu$ sec  
to 10 msec, and a pulse amplitude range of about 1 KV/cm  
to 20 KV/cm.

35

1           6. . The method of claim 5 wherein the pulses are  
selected from the group consisting of single pulses, train  
of pulses and multiple train of pulses.

5           7. The method of Claim 1, wherein said pulsed  
radiofrequency oscillating field includes a frequency of  
about 0.02 to 10 MHz; a pulse width of about 20 to 2000  
µsec and a pulse amplitude of 2-10 KV/cm.

10          8. The method of Claim 1, wherein said pulsed  
radiofrequency oscillating field includes different wave  
forms.

15          9. A method for fusing biological particles,  
comprising the steps of:  
          placing a plurality of biological particles in  
          solution between electrodes;  
          positioning said particles in close proximity; and  
20          fusing said biological particles by applying a  
pulsed radiofrequency oscillating electrical field  
across said electrodes.

25          10. The method of Claim 9, wherein said  
positioning step includes placing suspended biological  
particles in a container which allows said biological  
particles to congregate.

30          11. The method of Claim 9, wherein said  
positioning step includes:  
          applying an alternating electric field across  
          said electrodes for bringing said particles into close  
proximity.

35

1

12. The method of Claim 11, wherein said applying an alternating electrical field occurs before and after said fusing step.

5

13. The method of Claim 9, wherein said biological particles include a target cell and a particle preloaded with chemical agents or molecules for  
10 introducing chemical agents and molecules into cells.

14. The method of Claim 13, wherein:  
said particle preloaded with said chemical agents  
or molecules is selected from the group consisting of  
15 erythrocyte ghosts, liposomes, vesicles, isolated  
cells and cultured cells; and  
said target cell is selected from the group  
consisting of animal cells, human cells, plant cells,  
bacteria and yeast.

20

15. The method of Claim 13, wherein said chemical agents and molecules are selected from the group consisting of antibodies, proteins, drugs, molecular probes, hormones, growth factors, DNA, RNA, enzymes,  
25 organic chemicals and inorganic chemicals.

30

16. The method of Claim 9, wherein said biological particles include cells from at least two different species.

35

17. The method of Claim 9, wherein said biological particles include an antibody producing cell and a continuously dividing cancer cell.

1  
18. The method of Claim 9, wherein said  
biological particles include hyperimmunized mouse spleen  
cells and myeloma cells.

5  
19. The method of claim 9, wherein said  
biological particles include human lymphocytes and human  
cancer cells.

10  
20. The method of Claim 9, wherein said  
biological particles include cells of at least two  
different types.

21. A method for fusing biological particles,  
15 comprising the steps of:

placing a plurality of biological particles in  
solution between electrodes, wherein said biological  
particles are selected from the group consisting of  
animal cells, human cells, plant cells, vesicles,  
20 erythrocyte ghosts, liposomes, protoplasts, bacteria  
and yeasts;

providing a low amplitude alternating electrical  
field across said electrodes for bringing said  
particles into close proximity by dielectrophoresis,  
25 wherein said electrical field has a frequency range of  
about 60 Hz to about 10 mega Hz and a field strength  
of about 100 to 800 V/cm.;

applying a pulsed single or multi-frequency  
radiofrequency oscillating electrical field across  
said electrodes for fusing said particles, wherein  
30 said radiofrequency oscillating electrical field  
includes a frequency range of about 10 KHz to 100 MHz;  
a pulse width range of about 1  $\mu$ sec to 10 msec, and  
a pulse amplitude range of about 1 KV/cm to 20 KV/cm;  
35 and

1 providing an alternating electrical field across  
said electrodes for bringing said particles into close  
proximity for fusion, wherein said electrical field  
5 has a frequency range of about 60 Hz to about 10 mega  
Hz and a field strength of about 100 V/cm to 800 V/cm.

22. As a composition of matter, a hybridoma cell  
made by a method for fusing antibody producing cells with  
10 continuously dividing cancer cells, comprising the steps  
of:

suspending antibody producing cells and cancer  
cells in solution between electrodes;  
providing an alternating electrical field wherein  
15 said electric field forms pearl chains by allowing the  
cells to move dielectrophoretically; and  
forming said hybridoma cell by applying a high  
strength pulsed radiofrequency oscillating field  
across said electrodes.

20 23. The hybridoma cell of Claim 22, wherein said  
antibody producing cells are hyperimmunized mouse spleen  
cells and the cancer cells are myeloma cells.

25 24. The hybridoma cell of claim 22, wherein said  
antibody producing cells are human lymphocytes and the  
cancer cells are human cell lines.

25 25. A method of inserting foreign genes into  
biological particles comprising the steps of:

30 placing a plurality of biological particles  
between electrodes in a solution containing the  
foreign gene; and  
applying a pulsed radiofrequency oscillating  
35 electrical field across said electrodes.



1           26. The method of claim 25, wherein,  
            said pulsed radiofrequency includes a frequency  
            range of about 10 KHz to 10 MHz, a pulse width range  
5           of about 1  $\mu$ sec to 10 msec and a pulse amplitude of  
            about 1 to 20 KV/cm;

            said biological particles are selected from the  
            group consisting of human cells, animal cells, plant  
            cells, protoplasts, bacteria and yeasts.

10           27. A method of treating genetic diseases,  
            comprising the steps of:

            inserting in vitro a gene into cells by the  
15           method of claim 25; and

            transplanting said cells with the inserted gene  
            into the organism with the genetic defect.

20           28. The method of claim 27, wherein the organism  
            is a human.

            29. The method of claim 27, wherein, the cells  
            in which the gene is inserted are extracted from the  
            organism with the genetic defect prior to the insertion  
25           steps.

            30. The method of claim 27, wherein, the genetic  
            disease is sickle cell anemia, the cells are bone marrow  
            stem cells and the gene is the normal hemoglobin gene.

30           31. The method of claim 27, wherein,  
            the genetic disease is selected from the group  
            consisting of hemophilia, diabetes,  
            hypercholesterolemia and growth disorders;

35

1           the cells are selected from the group consisting  
of bone marrow stem cells, pancreatic islet cells,  
liver cells and other human cells; and

5           the gene is selected from the group consisting of  
the normal clotting factor VIII gene, the normal  
insulin gene the normal low density lipoprotein  
receptor gene and the growth hormone gene.

10           32. The method of Claim 27, wherein said cells  
are bone marrow stem cells.

33. A device for the poration and fusion of  
biological particles comprising:

15           a container, including a non-conducting material  
capable of holding liquid, wherein said container  
includes an access port for receiving said biological  
particles;

20           electrodes positioned equidistant from each other  
in said container; and

          a high power function generator attached to said  
electrodes for applying an electrical field including  
a pulsed radiofrequency electric field.

25           34. The device of Claim 33, wherein said  
container has a shape which allows the biological  
particles to congregate by gravity.

30           35. The device of Claim 33, wherein said  
radiofrequency is a pulsed radiofrequency oscillating  
field and includes a frequency range of about 10 KHz to  
100 MHz; a pulse width range of about 1  $\mu$ sec to 10 msec;  
and a pulse amplitude range of about 1 KV/cm to 20 KV/cm.

35           36. The device of Claim 33, wherein said  
function generator further includes the capability of

1 applying a continuous low power alternating current for  
bringing said biological particles into close proximity  
for fusion.

5 37. The device of Claim 36, wherein said low  
power alternating current includes a frequency range of  
about 60 Hz to the 10 mega Hz range and a field strength  
of about 100 V/cm to 800 V/cm.

10 38. A device for optical microscopic observation  
of poration and fusion of biological particles comprising:  
a container with transparent bottom;  
electrodes inserted into said container, wherein  
15 said electrodes are equidistant from each other and  
arranged in a pattern which allows at least ten  
microliter of cells to closely contact the electrodes;  
and

20 a high power function generator attached to said  
electrodes for applying an electrical field including  
radiofrequency.

39. A device for cell poration and fusion of  
biological particles, comprising:

25 a handle of non-conductive material for  
manipulating said device;  
electrodes positioned equidistant from each other  
and attached to said handle; and  
a connecting means for attaching the electrodes  
30 to a high power function generator.

40. The device of Claim 39, wherein said  
electrodes are wrapped around a non-conducting core in the  
form of a double helix.

1           41. The device of Claim 39, wherein said  
electrodes comprise:  
            segmented shapes; and  
5           insulating spacers for separating said segmented  
            shapes.

            42. The device of Claim 39, wherein said  
electrodes are bottom-contact electrodes selected from the  
10           group consisting of spirals, concentric rings, concentric  
            squares, parallel plates and interdigitating arrays.

            43. The device of claim 39, wherein said  
electrodes are coaxial and are about 0.5 to 2.0 mm apart,  
15           have a length of about 0.5 to 3 cm and a diameter  
            sufficiently small to fit into a well of a multi-well  
            culture plate.

            44. A high power function generator, comprising:  
20           a RF pulse generator including a gating circuit  
            for gating the output of a radiofrequency oscillator  
            and a power amplifier for generating the high power RF  
            pulse from the gated output of the radiofrequency  
            oscillator;  
25           an AC field generator; and  
            a mercury wetted relay for switching between the  
            RF pulse and the AC field.

            45. A device for cell poration and cell fusion,  
30           comprising:  
            a digital computer for synthesizing and  
            generating a RF wave form and an AC wave form;  
            a amplifier to convert the wave form generated by  
            the digital computer to high power wave forms; and  
35           an electrode communicating with said

1           amplifier for applying the electric field to  
            biological cells.

5           46. The device of claim 45 further including an  
            information storage device.

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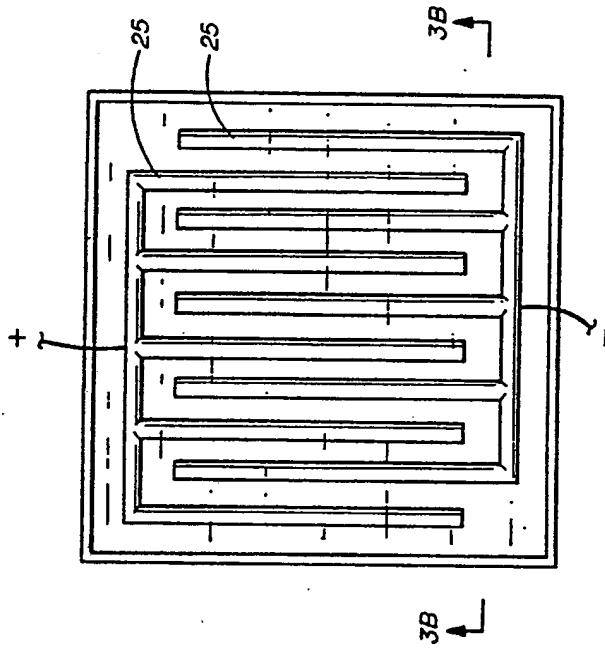


FIG. 3A

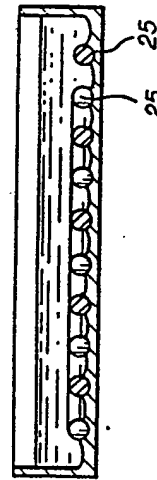


FIG. 3B

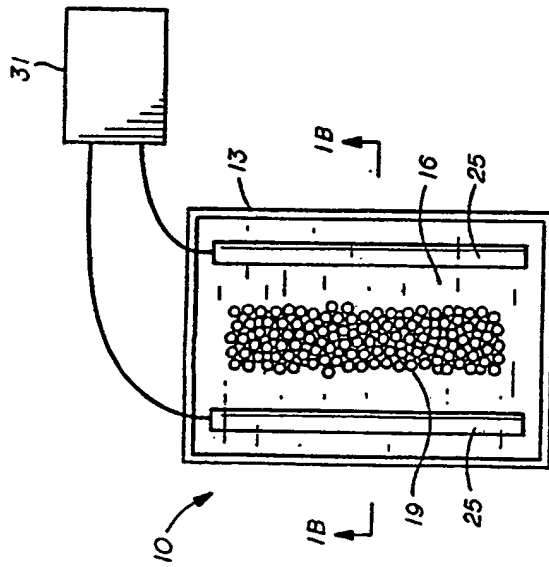


FIG. 1A

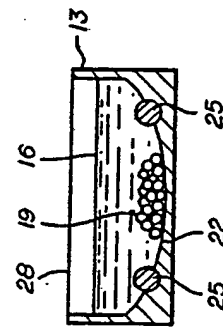
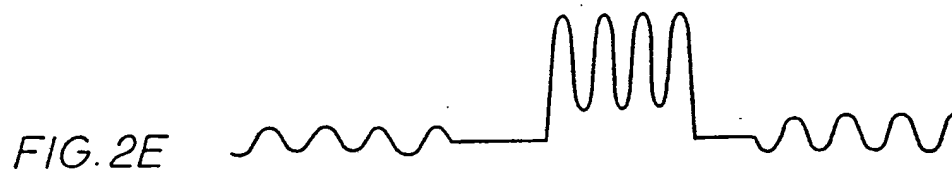
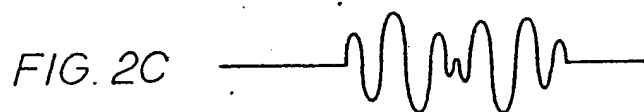
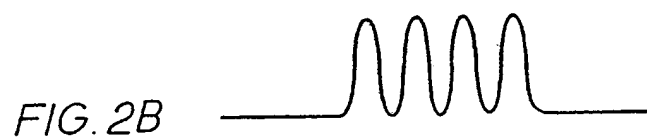
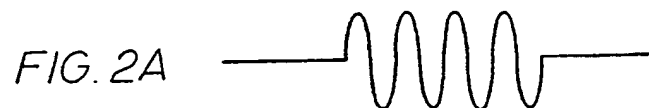


FIG. 1B

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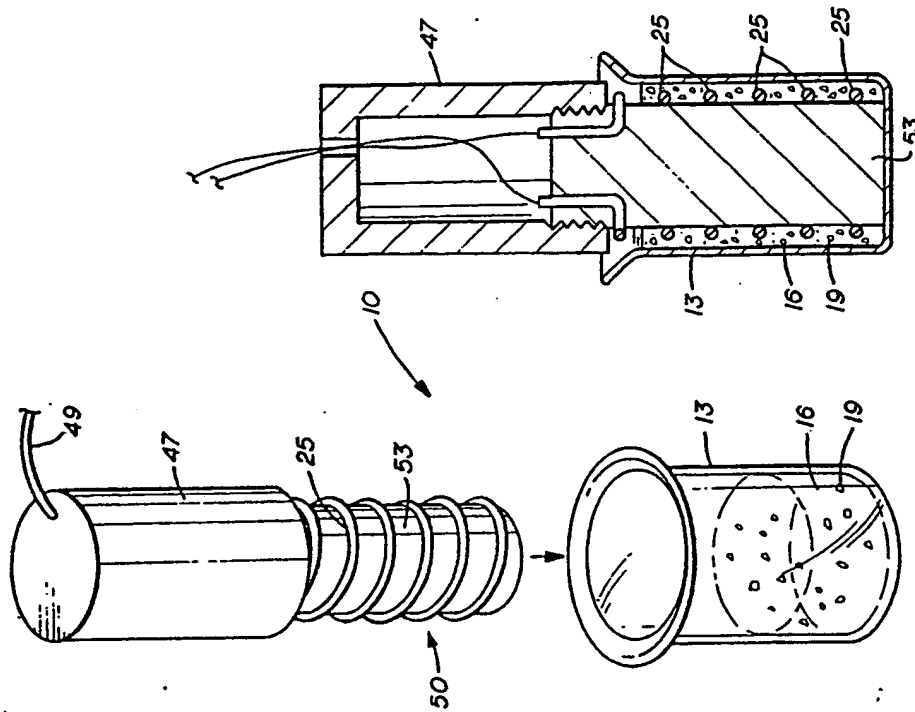


FIG. 5B

FIG. 5A

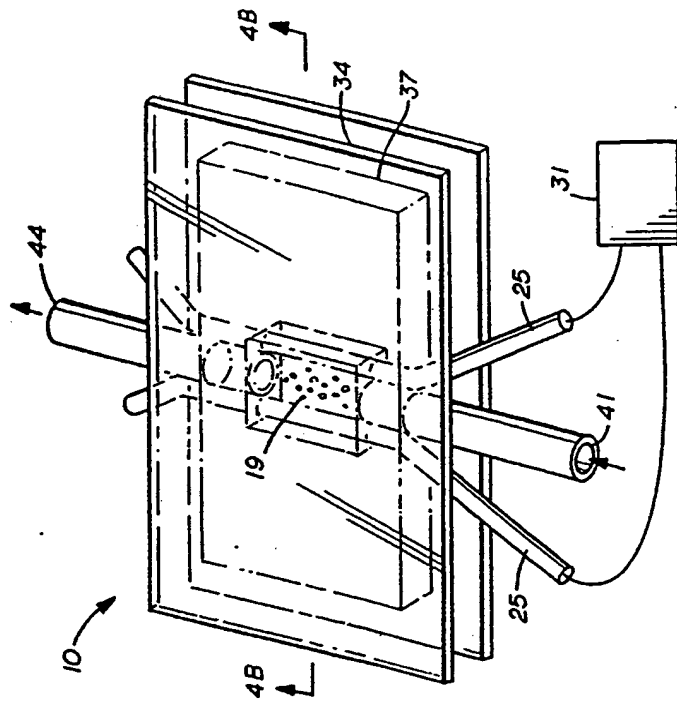


FIG. 4A

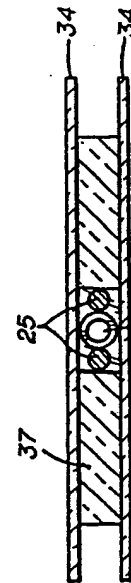


FIG. 4B



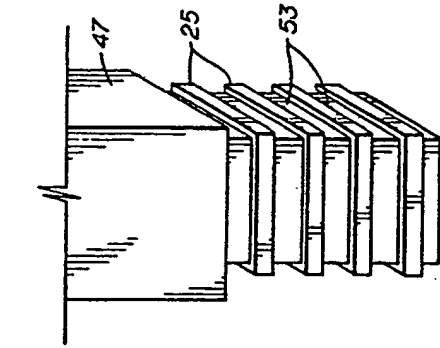


FIG. 8A

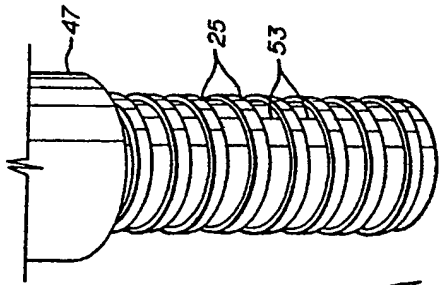


FIG. 7A

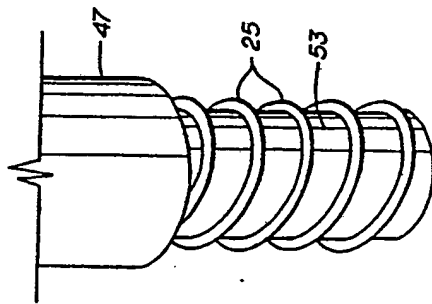


FIG. 6A

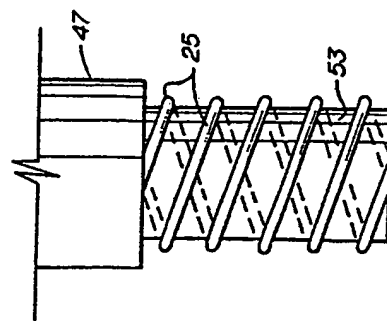


FIG. 6B

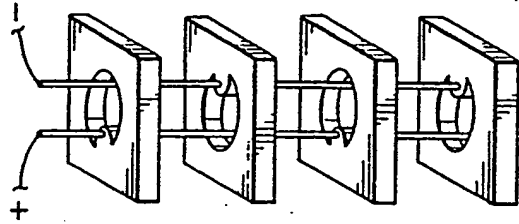


FIG. 8B

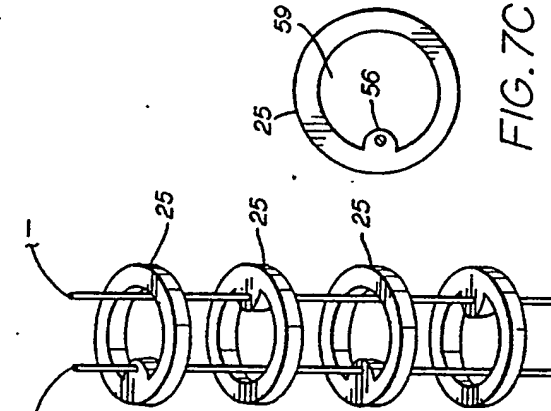


FIG. 7B

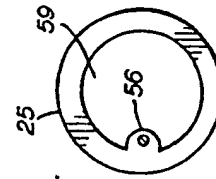


FIG. 7C

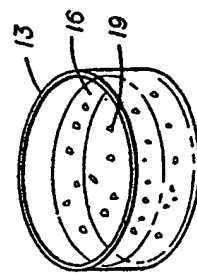
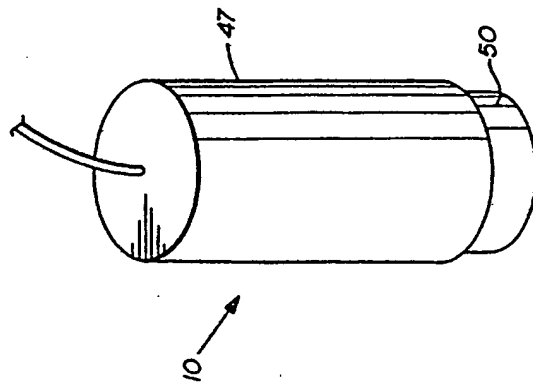


FIG. 9

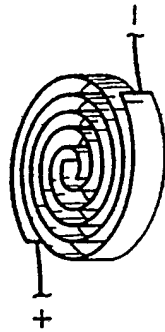


FIG. 10A



FIG. 10B

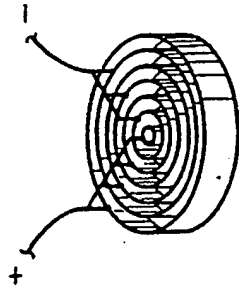


FIG. 11A

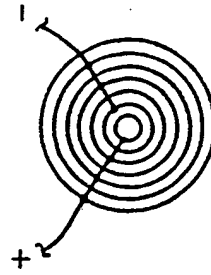


FIG. 11B

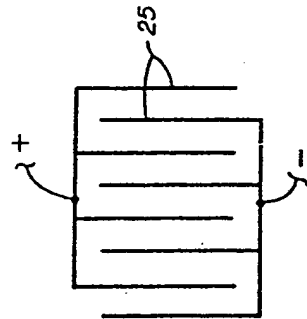


FIG. 12C

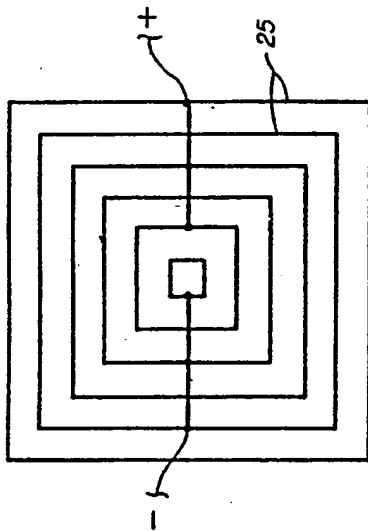


FIG. 12B

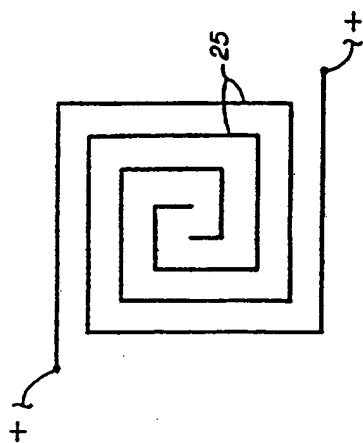


FIG. 12A

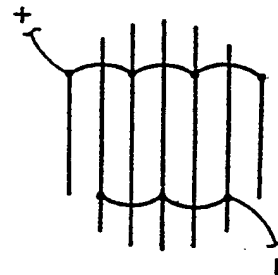


FIG. 12D

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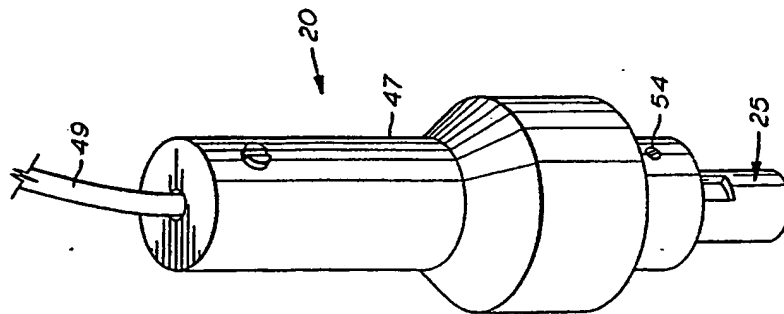


FIG. 13A

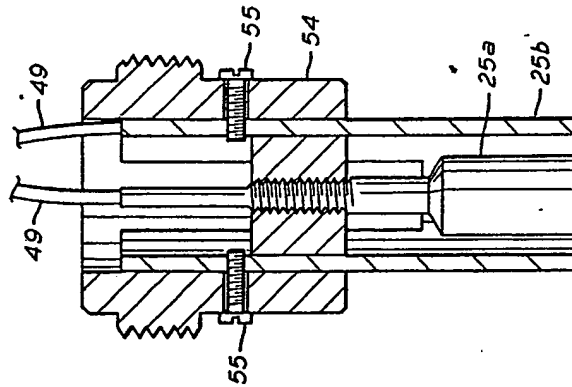


FIG. 13B

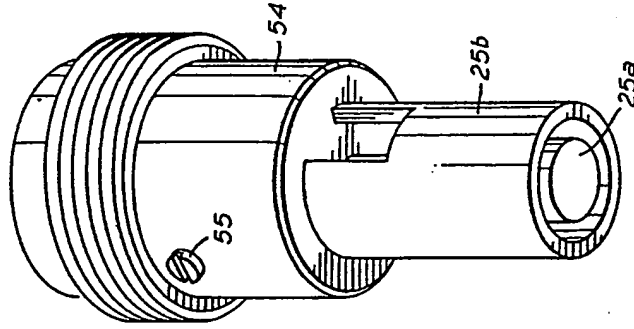


FIG. 13C

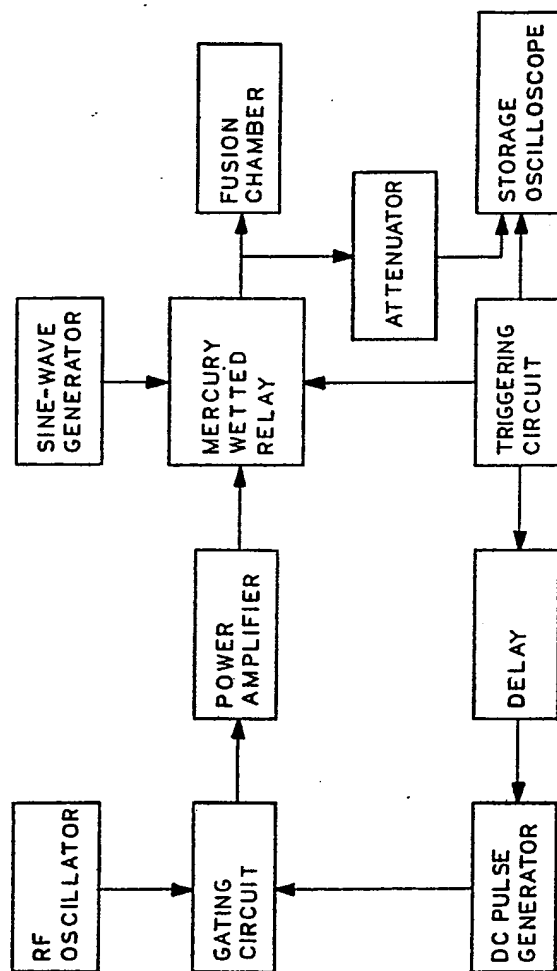


FIG. 14

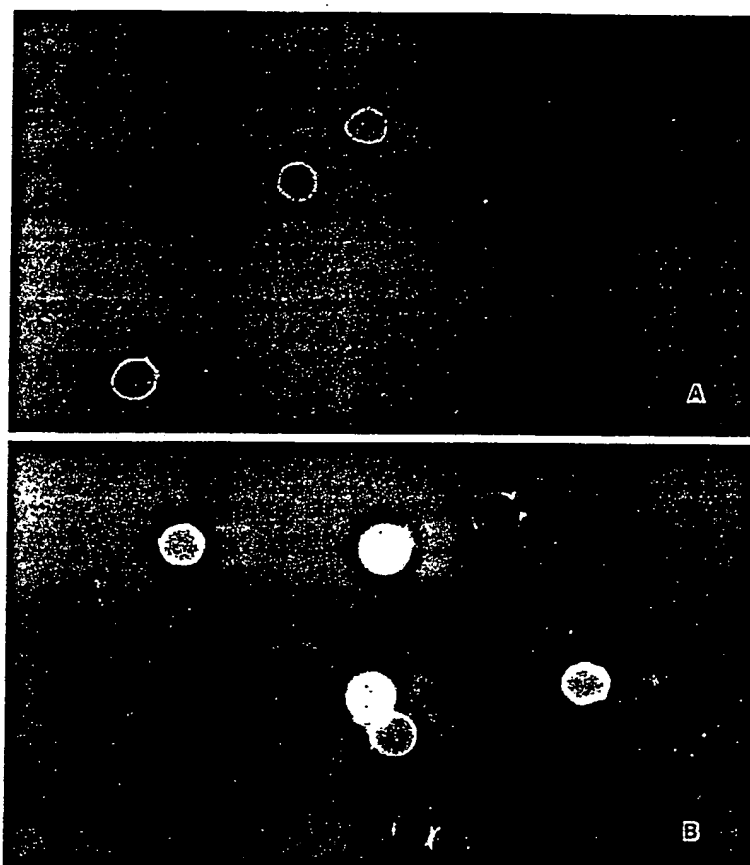


FIG. 15

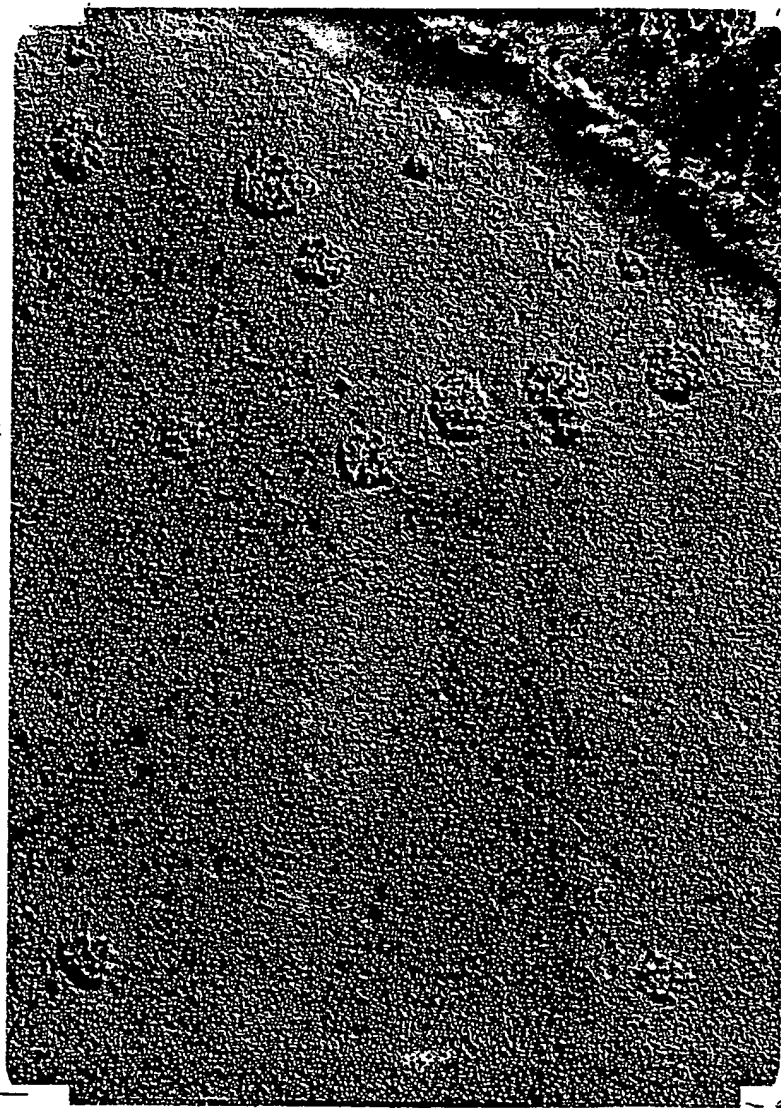


FIG. 16

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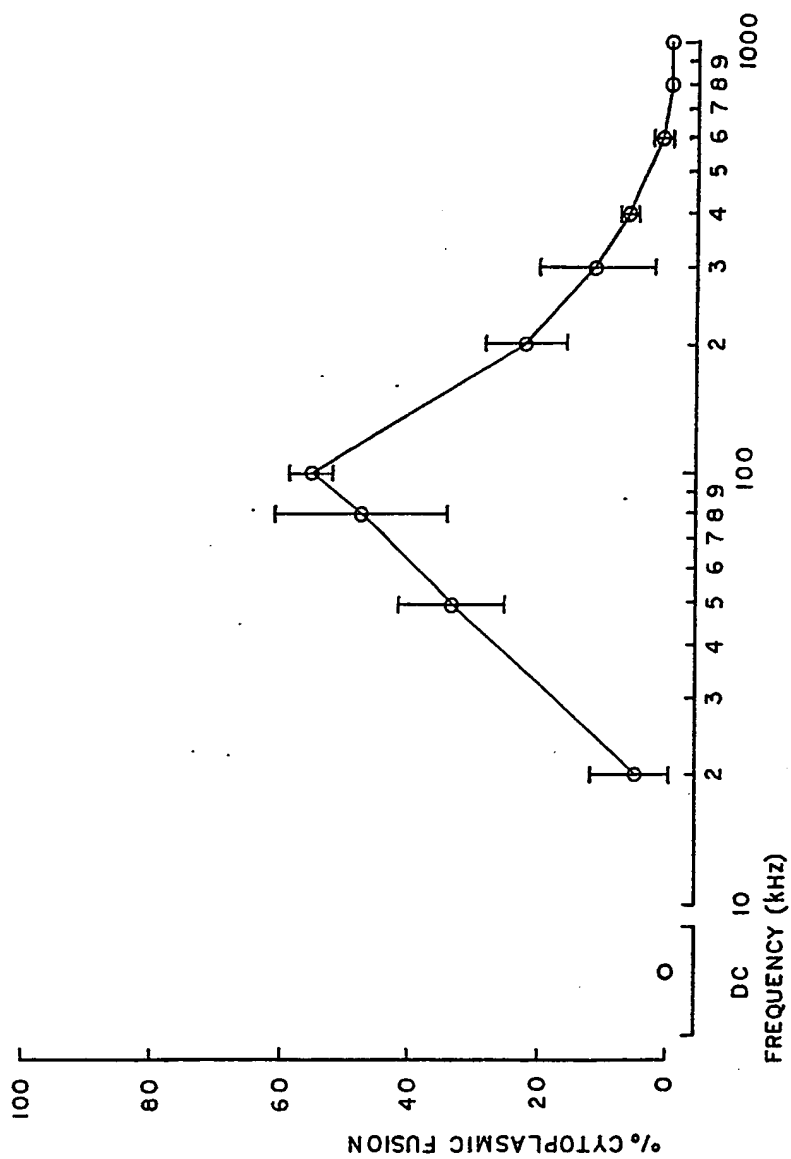


FIG. 17



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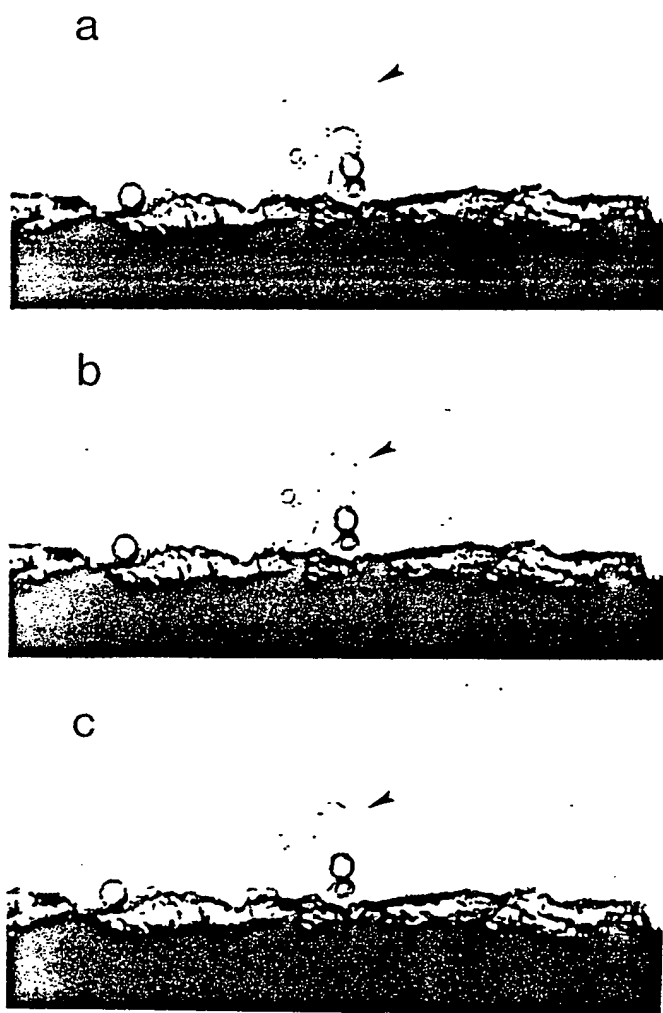


FIG. 18



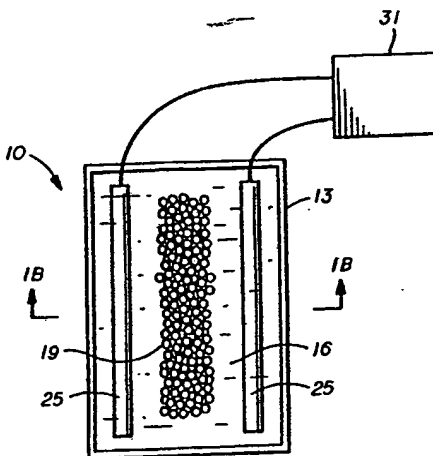
INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification <sup>4</sup> :  C12N 15/00, 13/00		A3	(11) International Publication Number: WO 89/ 03426
			(43) International Publication Date: 20 April 1989 (20.04.89)
(21) International Application Number: PCT/US88/03457		(81) Designated States: AT (European patent), AU, BE (European patent), CH (European patent), DE (European patent), FR (European patent), GB (European patent), IT (European patent), JP, LU (European patent), NL (European patent), SE (European patent).	
(22) International Filing Date: 5 October 1988 (05.10.88)			
(31) Priority Application Numbers: 106,282 238,607		Published	
(32) Priority Dates: 9 October 1987 (09.10.87) 30 August 1988 (30.08.88)		<i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>	
(33) Priority Country: US		(88) Date of publication of the international search report: 13 July 1989 (13.07.89)	
(71) Applicant: BAYLOR COLLEGE OF MEDICINE [US/US]; One Baylor Plaza, Houston, TX 77030 (US).			
(72) Inventor: CHANG, Donald, C. ; 6306 Coachwood, Houston, TX 77035 (US).			
(74) Agent: PAUL, Thomas, D.; Fulbright & Jaworski, 1301 McKinney St., #5100, Houston, TX 77010 (US).			

**(54) Title: METHOD OF AND APPARATUS FOR CELL PORATION AND CELL FUSION USING RADIOFREQUENCY ELECTRICAL PULSES**

**(57) Abstract**

Disclosed are an apparatus and a method for the poration and fusion of cells using high-power radiofrequency electrical pulses. The electrodes of the apparatus can be hand held or part of integrated equipment with special containers for cells. The electrodes, which are positioned equidistant from each other, are attached to a high power function generator. The power function generator can apply a continuous AC electrical field and/or a high-power pulsed radiofrequency electrical field across the electrodes. The alternating electrical field induces cell congregation by the process of dielectrophoresis. The high-power pulsed radiofrequency electrical field porates or fuses the cells.



The method has the ability to porate or fuse biological cells with a very high efficiency. The method can be used to fuse or porate a variety of cells including animal cells, human cells, plant cells, protoplasts, erythrocyte ghosts, liposomes, vesicles, bacteria and yeasts. The method has the unique ability to porate or fuse cells in very small or very large numbers. During the poration or fusions, a variety of chemical agents including DNA, RNA, antibodies, proteins, drugs, molecular probes, hormones, growth factors, enzymes, organic chemicals and inorganic chemicals can be introduced into these cells. The method can also be used to produce new biological species, to make hybridoma cells which produce animal or human monoclonal antibodies and to insert therapeutic genes into human cells which can be transplanted back into the human body to cure genetic diseases.

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FI	Finland				

# INTERNATIONAL SEARCH REPORT

International Application No PCT/US 88/03457

<b>I. CLASSIFICATION OF SUBJECT MATTER</b> (if several classification symbols apply, indicate all) <sup>6</sup>		
According to International Patent Classification (IPC) or to both National Classification and IPC		
IPC <sup>4</sup> : C 12 N 15/00; C 12 N 13/00		
<b>II. FIELDS SEARCHED</b>		
Minimum Documentation Searched <sup>7</sup>		
Classification System <sup>1</sup>	Classification Symbols	
IPC <sup>4</sup>	C 12 N	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched <sup>8</sup>		
<b>III. DOCUMENTS CONSIDERED TO BE RELEVANT<sup>9</sup></b>		
Category <sup>10</sup>	Citation of Document, <sup>11</sup> with indication, where appropriate, of the relevant passages <sup>12</sup>	Relevant to Claim No. <sup>13</sup>
A	EP, A, 0128567 (KERNFORSCHUNGSANLAGE JÜLICH GmbH) 19 December 1984, see claims --	1-4,11
A	US, A, 4578168 (G.A. NOFMANN) 25 March 1986, see claims; column 5 --	1
A	US; A, 3059359 (J.H. HELLER) 25 June 1963, see claims ----	1,5
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<b>IV. CERTIFICATION</b>		
Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report	
30th May 1989	19 JUN 1989	
International Searching Authority	Signature of Authorized Officer	
EUROPEAN PATENT OFFICE	P.C.G. VAN DER PINTEN	

**ANNEX TO THE INTERNATIONAL SEARCH REPORT  
ON INTERNATIONAL PATENT APPLICATION NO.**

US 8803457  
SA 25321

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Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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